

To prepare slides for microscopic examination, the method described by Meredith⁹ for mammalian tissue was used. The pieces of fixed hydra tissue were transferred into a Dreyer tube containing 0.5 ml 60% acetic acid. Within 5 min the tissue loses its coherence, and a cell suspension could be formed by aspiration with a finely-drawn pasteur pipette. A drop of cell suspension was then transferred onto a microscope slide on a hot plate at about 60°C and immediately withdrawn. The same drop of cell suspension was applied to the slide in this manner 5–10 times before discarding. The procedure was repeated with further drops of suspension until all the fixed material was applied onto the slide. The slides were then placed vertically for 10 min in a 5% aqueous solution of Giemsa stain, rinsed with tap water and air-dried.

The preparations were scanned for metaphases under the microscope. Chromosome counts were made for all metaphase spreads in which the chromosomes showed little or no overlap. We applied a preparation of three hydra per slide and could observe on average 12 metaphase spreads, of which an average of 2–3 spreads could be used for analysis.

Results. Chromosome counts were made from 31 spreads of *H. vulgaris* cells and 24 spreads of green hydra cells. Representative photomicrographs of spreads of each species are given in figure 1. For both species a range of chromosome counts were

obtained with a maximum of 34 chromosomes per spread, a minimum of 26, and most counts around 30 (fig. 2). The most likely explanations for the variation in chromosome counts between spreads are observational error and artifactual loss of chromosomes during preparation.

Discussion. A search of the literature shows that very few attempts have been made to develop a method for the examination of hydra chromosomes^{5,6,10–13} and those methods that have been developed involve cumbersome sectioning or squash procedures. The method described here should enable the researcher working with hydra to make chromosome counts easily and quickly. In this manner the species of hydra may be defined more precisely.

Previous reports on the chromosome number of *H. vulgaris* show $2n = 32$ (Niyama⁵; Datta⁶). Our counts are consistent with this number for the subspecies we used (*attenuata*). For green hydra no attempts to count the chromosome number has been reported since Dowing^{10,11}, who recorded $2n = 12$. Our counts of about 30 chromosomes per cell differ markedly from this number. It would be of much interest to verify whether the chromosome number does vary between species or strains of green hydra.

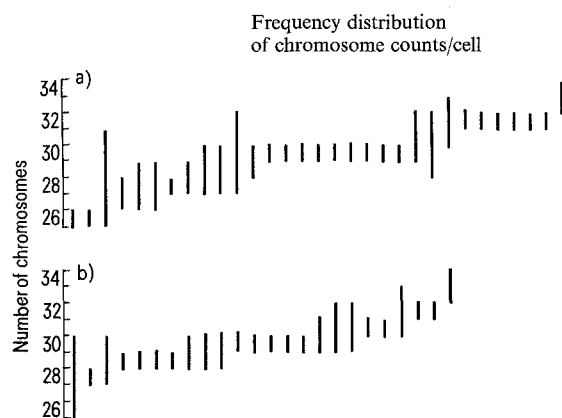


Figure 2. Chromosome counts based on spreads of *H. vulgaris* cells (a) and green hydra cells (b). Each bar represents one spread. For some spreads more than one interpretation in terms of chromosome number was possible.

- 1 This work was done while A.R. and M.R. were on leave in Oxford. We thank Prof. D.C. Smith F.R.S. for providing laboratory facilities. We wish to thank Prof. Smith and Dr A.E. Douglas for reading earlier drafts of this paper.
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Announcements

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Plant flavonoids in biology and medicine: biochemical, pharmacological, and structure-activity relationships

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This interdisciplinary symposium will review the role of flavonoids in plants and animals and their effects in numerous mammalian cells systems. Invited lecturers will review recent advances in our understanding of their significance in physiology and function and explore their potential therapeutic uses. For further information contact Dr Elliott Middleton, Jr, Department of Medicine, Buffalo General Hospital, Buffalo, N.Y. 14203/USA.

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10th European symposium on hormones and cell regulation
St.-Odile, Strasbourg, 30 September – 3 October 1985

Information can be obtained from Dr. R.J.B. King, Imperial Cancer Research Fund Laboratories, Hormone Biochemistry Department, P.O.Box 123, Lincoln's Inn Fields, London WC2A 3PX, England.